



# Detection of biological nitrification inhibition in canola

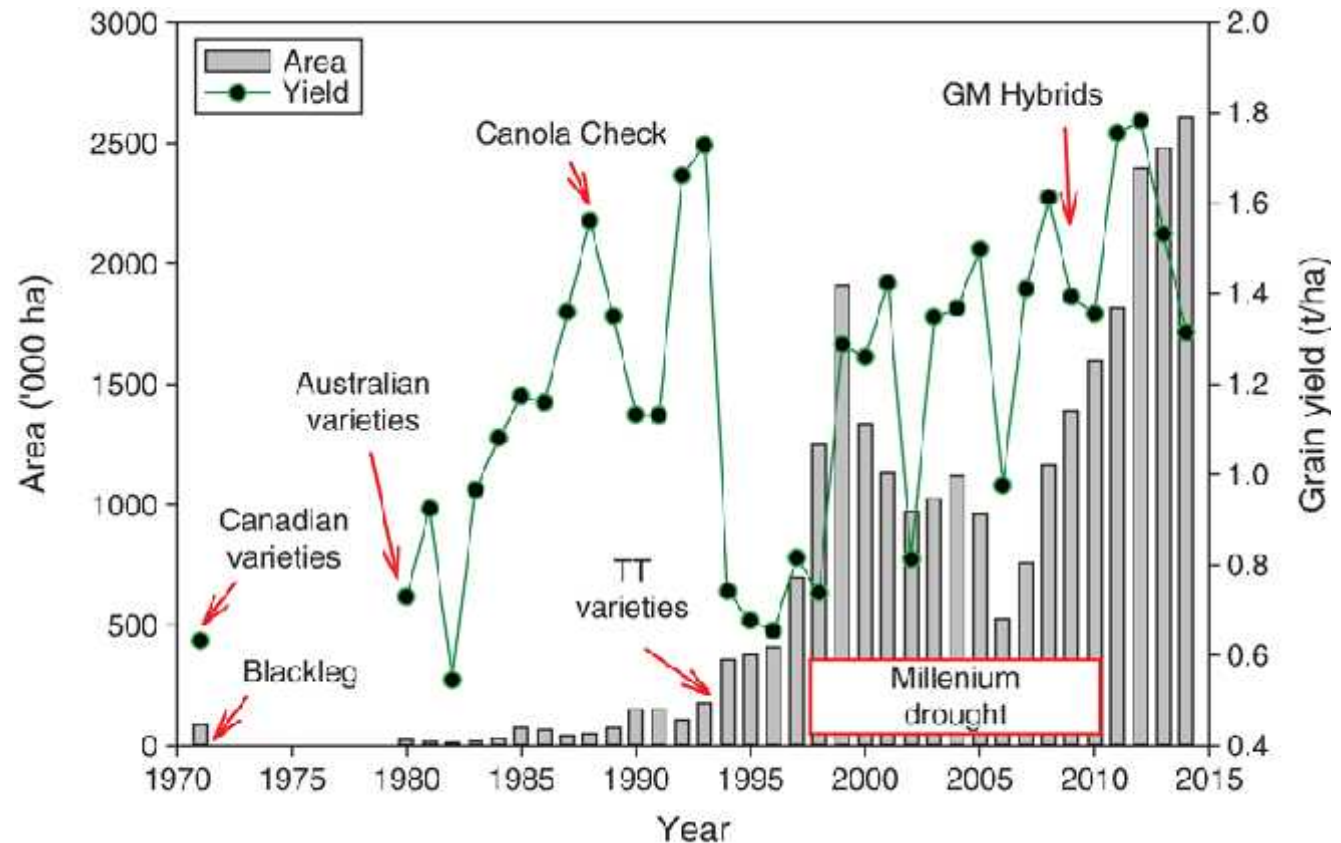
**Implications for N cycling and soil fertility in rotational cropping.**

Cathryn O'Sullivan, Elliott Duncan, Margaret Roper, Kelley Whisson, Karen Treble, Philip Ward

# Canola production is increasing in Australia

Growers have several reasons for planting canola

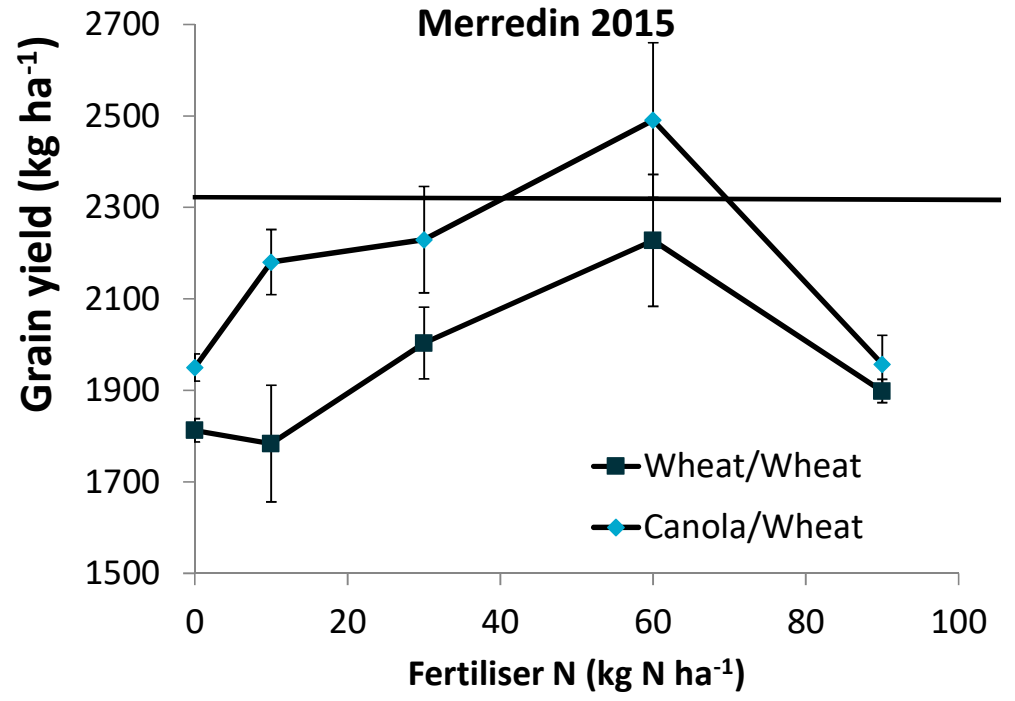
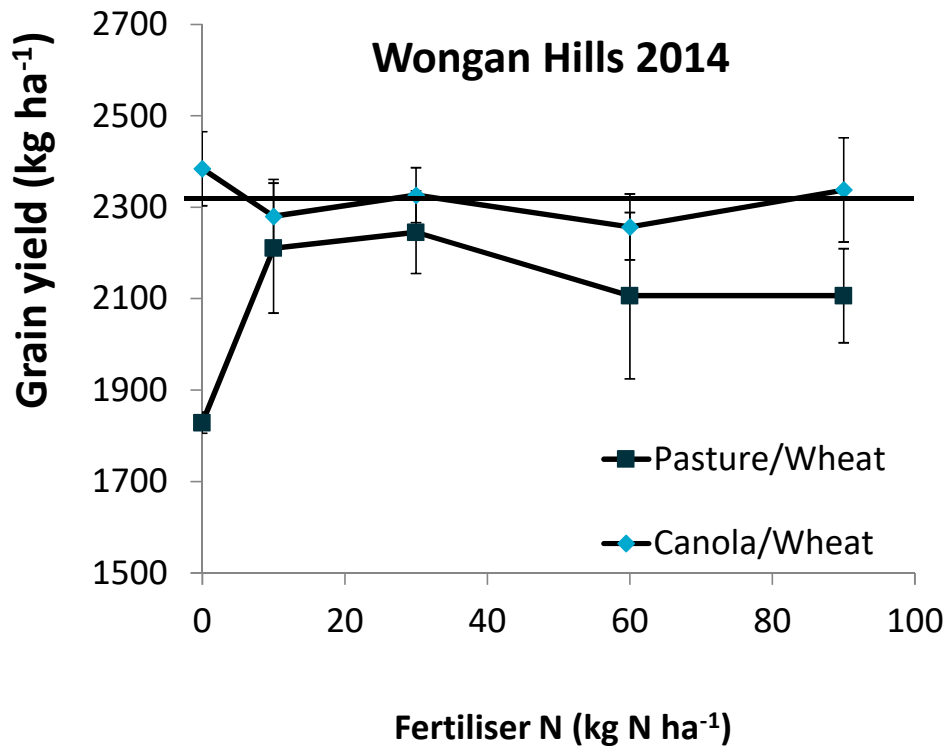
- Oilseed value
- Crop rotation, disease break
- Weed management



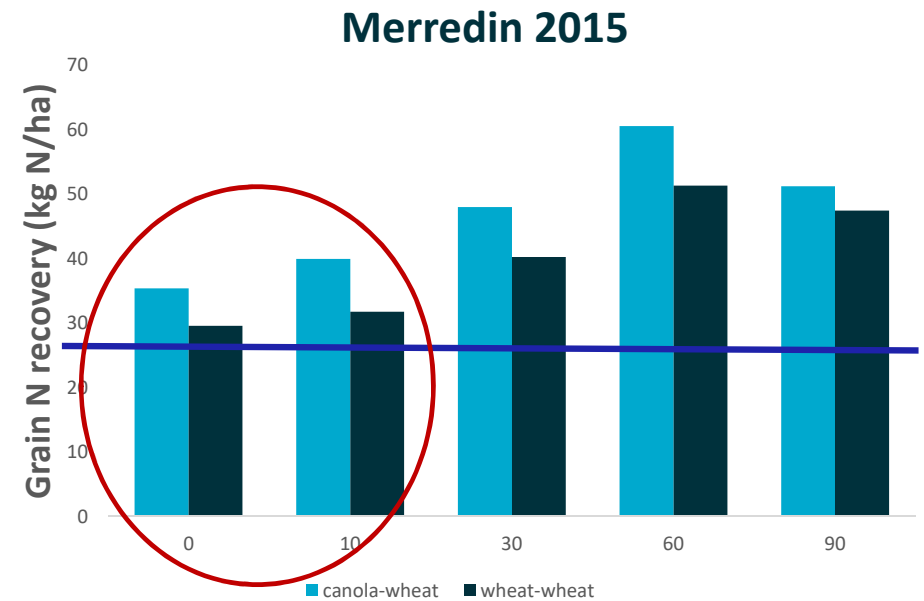
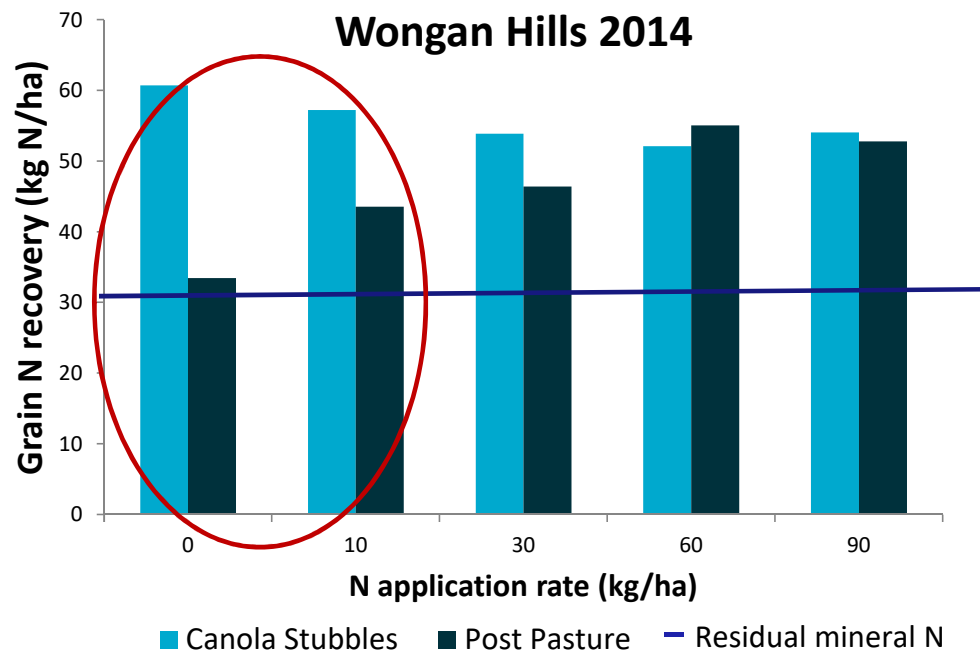
Kirkegaard, J. *et al.* Crop and Pasture Science 2016

# Increased N recovery following canola

## More than just the disease break?



# N recovery greater than residual N even without fertiliser



# Research questions

Are Brassicas manipulating the N cycling in their rhizosphere?

Is there a store of organic N after a canola crop that serves as an N source for the following crop?

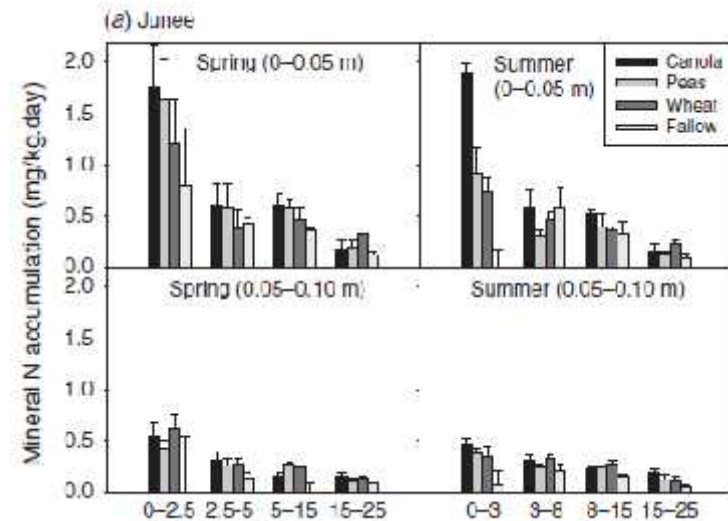
- Do brassicas produce root exudates with BNI?
- Does this slow nitrification in soils?
- What are the implications for other N cycling rates?
  - Immobilisation
  - Mineralisation



# Evidence of N retention after canola

**Mineral N accumulation** over the summer fallow **39–49 kg/ha higher** following brassicas compared with wheat at 2 sites

In lab incubations Brassica root tissues initially **immobilised**, and later released, mineral N at a **greater rate** than wheat root tissues.



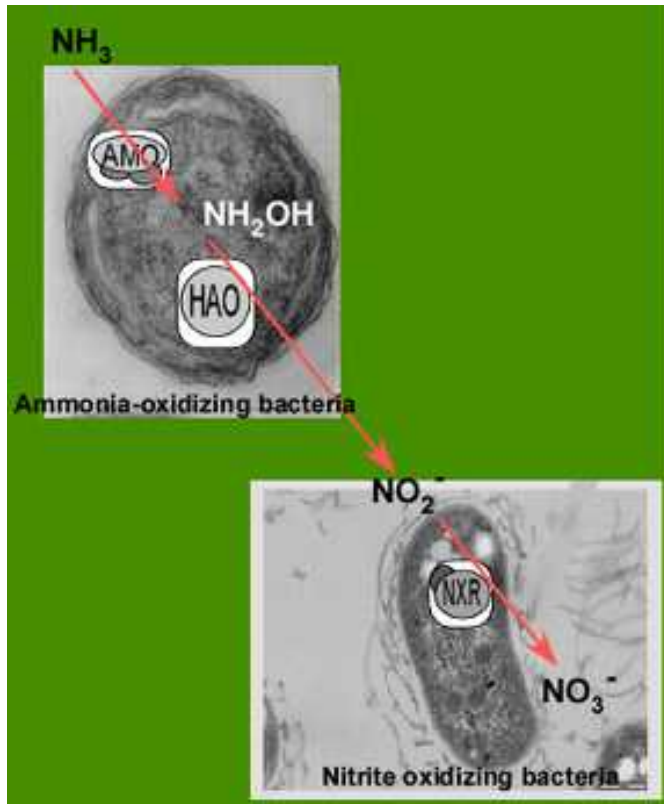
Ryan et al 2006

Accumulation of mineral-N following canola (94 kg/ha), **3 times as much as that following cereals**, and significantly higher than that after the legumes (50 kg/ha)

Sampling position	Previous crop	NH <sub>4</sub> <sup>+</sup> oxidisers <sup>D</sup>	NO <sub>2</sub> <sup>-</sup> oxidisers <sup>D</sup>
In row	Wheat	1.67	477
In-row	Canola	0.52	294
		***	n.s.
Between-row	Wheat	7.03	167
Between-row	Canola	0.66	729
		*	n.s.

Kirkegaard et al 1999

# Nitrification inhibition

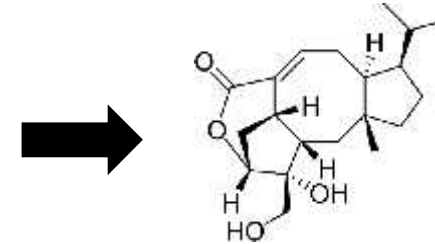


<http://nitrificationnetwork.org/gallery.php>

## Biological nitrification inhibitors



*Brachiaria humidicola*



brachialactone

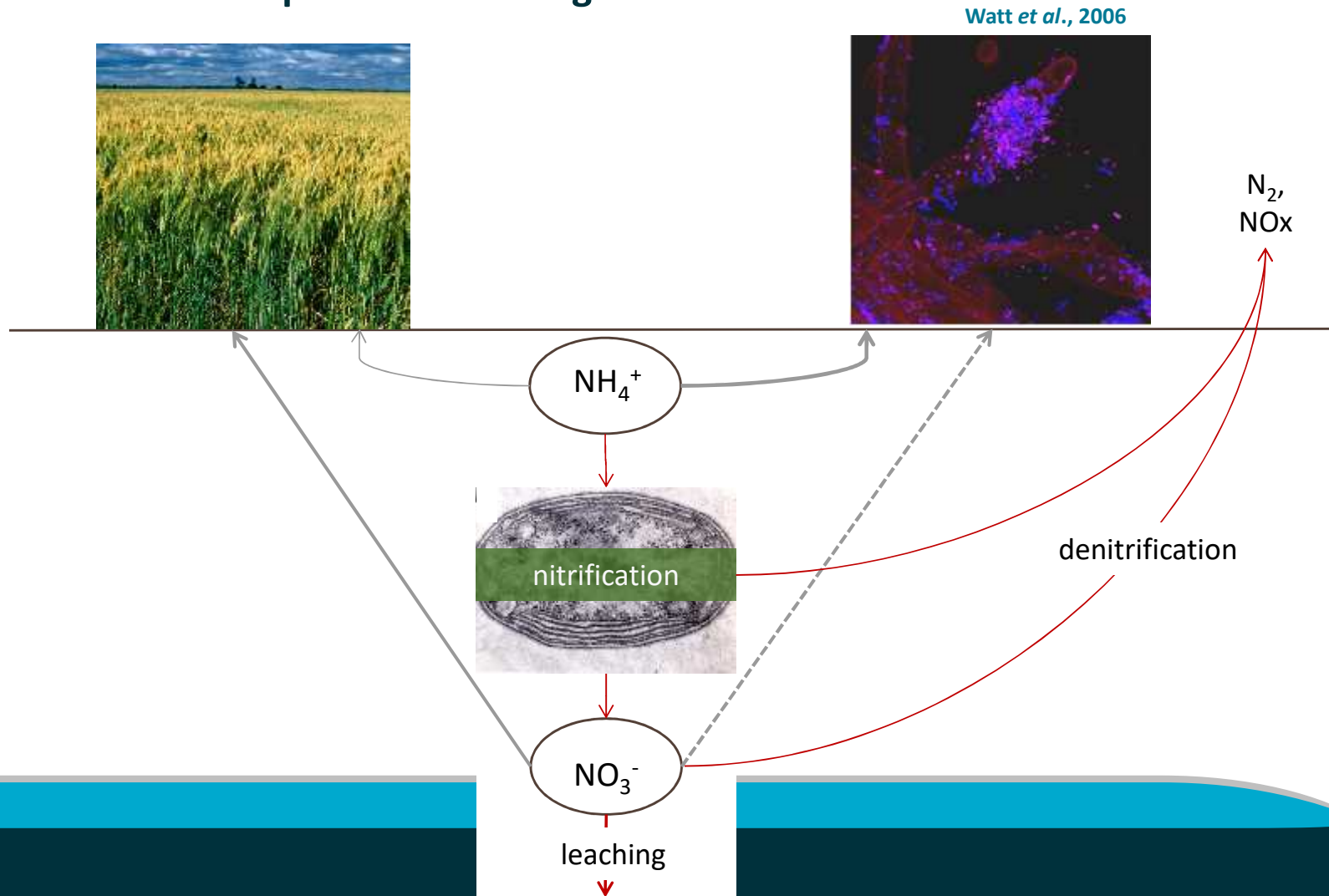
Subbarao et al 2009

- Formed by plant and released at root surface
- Continually replenished
- Found in *Brachiaria*, sorghum, pearl millet, peanut and *Leymus racemosus* (wild relative of wheat)

## Plants manipulating their microbiome

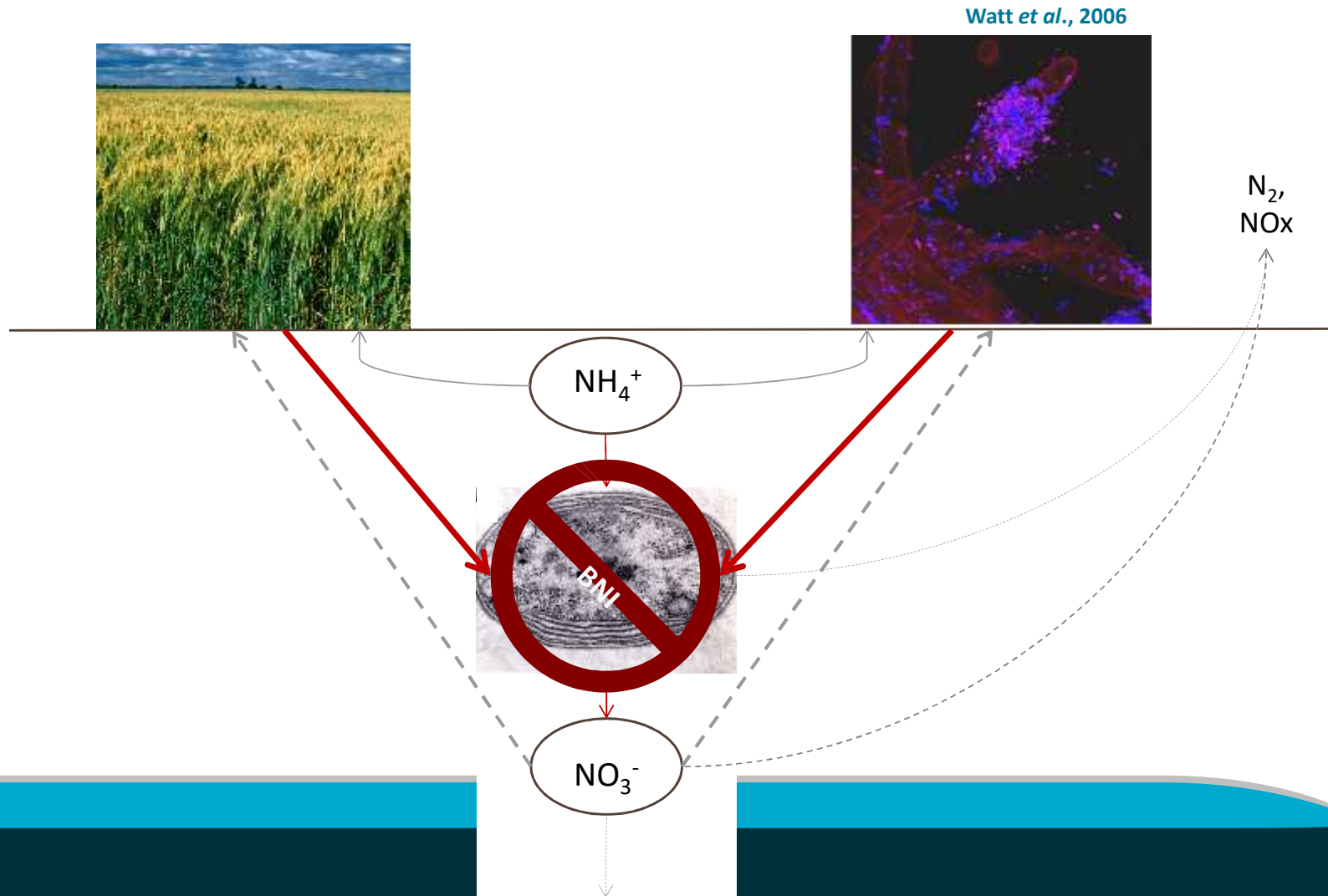
# BNI effects in crops and weeds

NUE improvement vs competitive advantage





# BNI effects on crops and microorganisms



# Assay to assess root extracts



Step 1  
Grow plants

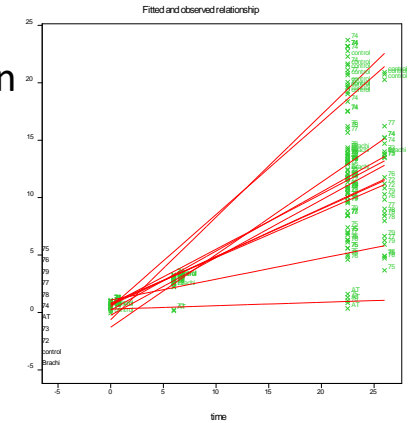


Step 2  
Extract compounds from roots

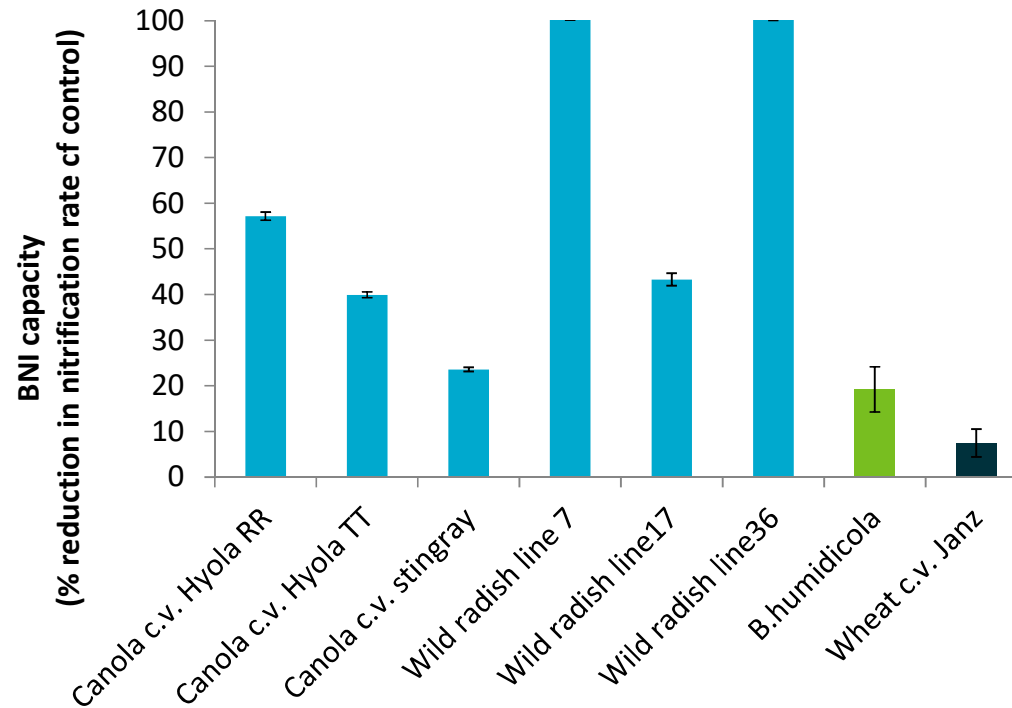


Step 4  
Assess nitrification rate against positive control (AT) and negative control (no inhibitor)

Step 3  
Grow pure cultures of *Nitrosomonas* and *Nitrosospira* in presence and absence of root extract



# BNI in root exudates



- BNI in canola and wild radish significantly higher than in B. humudicola
- First evidence of BNI in non-legume dicot.

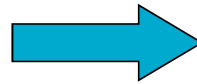
# Measuring BNI in pots



Step 1  
Grow  
plants



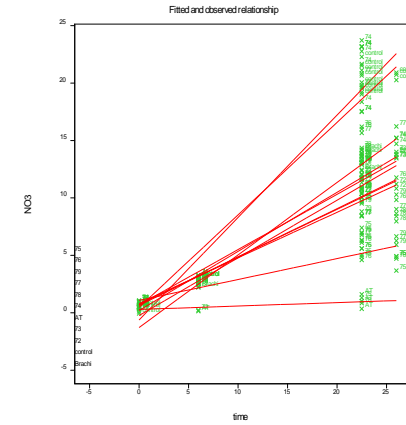
Step 2  
Remove plant  
and collect  
soil



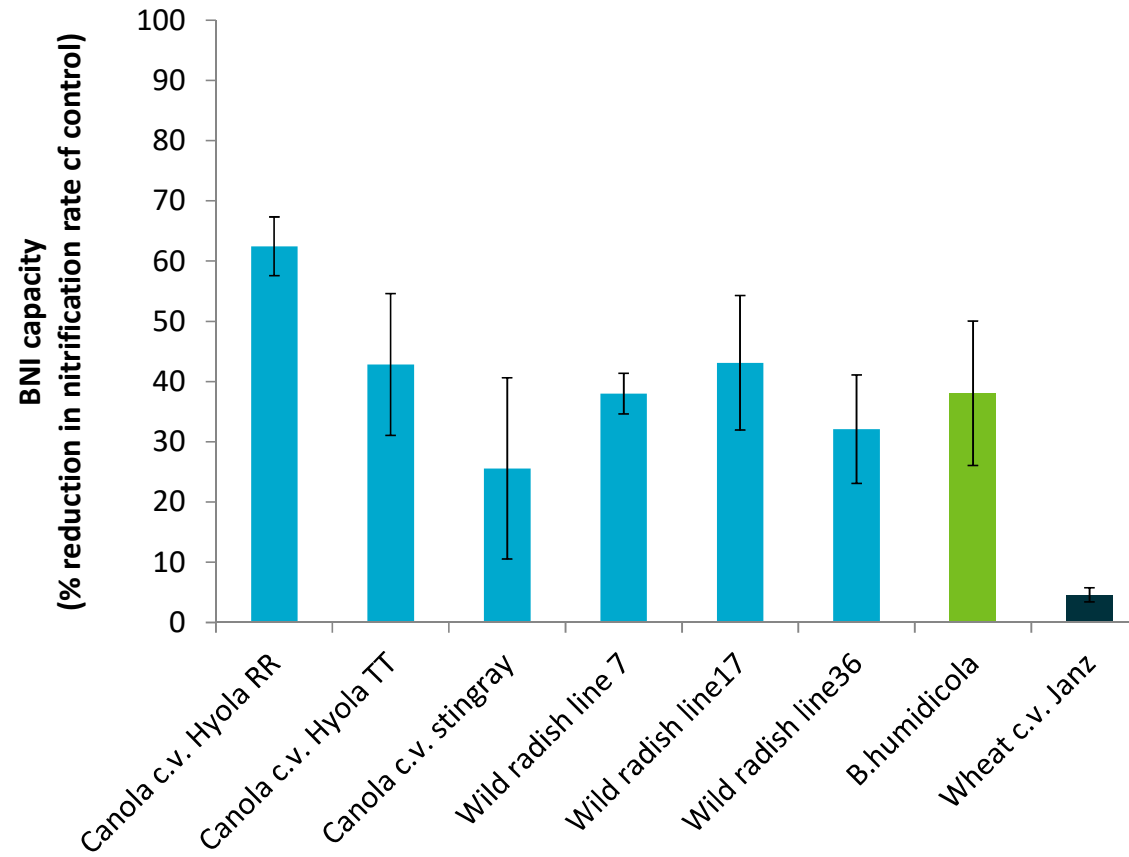
Step 4  
Assess  
nitrification  
rate against  
unplanted  
control



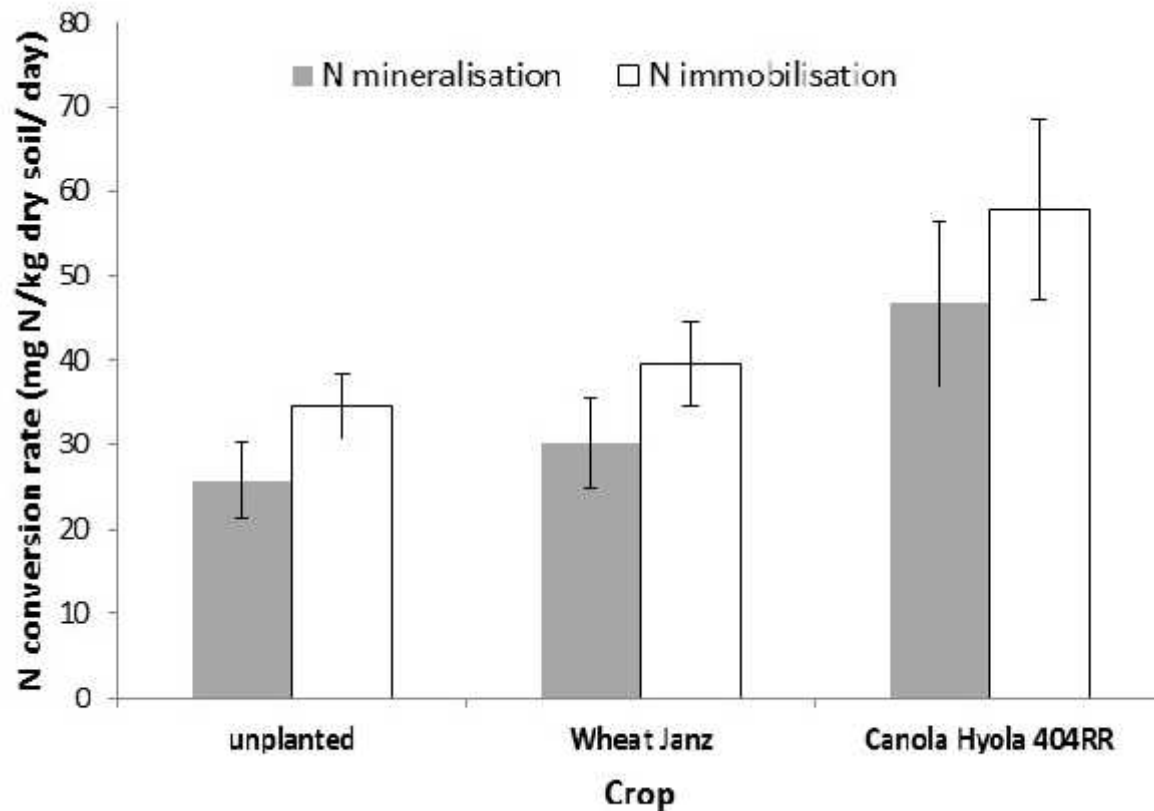
Step 3  
Incubate soil  
samples in  
slurry  
containing  
 $\text{NH}_4^+$



# BNI impact in pot assays



# 15N tracer soil incubations



## Net N Immobilisation in fallow following:

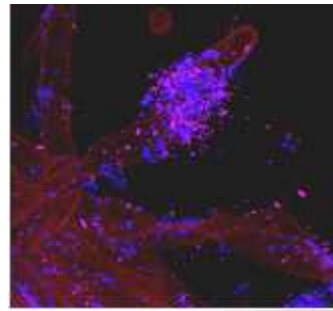
no plant – 8.8 mg N/kg soil.day

wheat - 9.3 mg N/kg soil.day

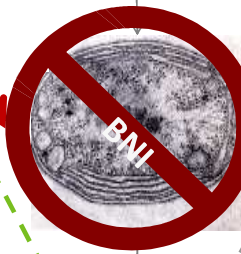
canola - 11.3 mg N/kg soil.day

# Conclusions

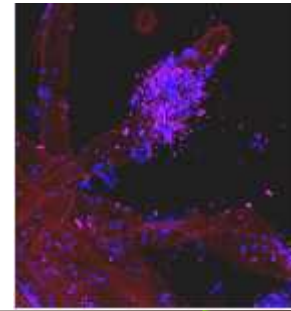
## Year 1 – canola rotation



$N_2$ ,  
 $NO_x$



## Year 2 – wheat rotation



$N_2$ ,  
 $NO_x$



leaching

denitrification

# Acknowledgements

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- DAFWA
- CSIRO Future Building Projects
  
- Project team
  - Elliott Duncan
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  - Kelley Whisson
  - Karen Treble
  - Phil Ward

Thank You